

Cite this: *RSC Advances*, 2011, 1, 573–575

www.rsc.org/advances

COMMUNICATION

Solid phase synthesis of ultra-photostable cyanine NIR dye library†

Raj Kumar Das,^a Animesh Samanta,^a Hyung-Ho Ha^b and Young-Tae Chang^{*ac}

Received 21st July 2011, Accepted 21st July 2011

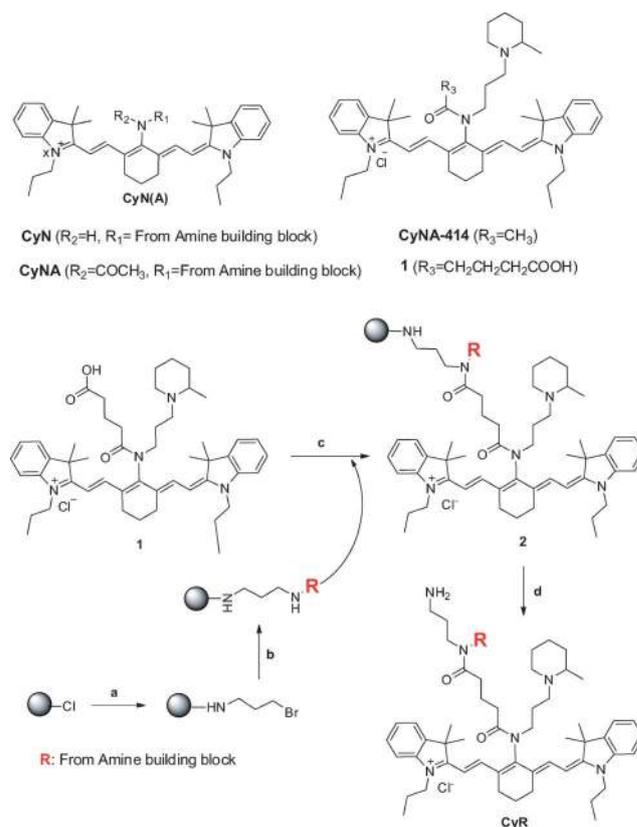
DOI: 10.1039/c1ra00498k

Photostability is one of the key issues in NIR dyes and we have previously reported a photostable CyNA library. Herein we report an ultra-photostable cyanine-based NIR fluorescence library, CyR, utilizing the stability component of CyNA. Efficient solid phase chemistry was also devised to provide a robust synthetic route to the new library.

Near-infrared (NIR) fluorescence (λ_{max} : 700–1000 nm) has recently received substantial attention in various chemical and biological studies.^{1–2} The advantages of NIR fluorescence are the deep penetrating ability through tissue and the significant reduction of autofluorescence which often occurs in visible light emission. Therefore, NIR probes are extensively used in a broad range of biological research as *in vivo* imaging probes,³ protein labelling agents⁴ and fluorescence tags in DNA sequencing.⁵ To date, a very limited number of scaffolds such as squaraine,⁶ quinone,⁷ triphenylmethane⁸ and cyanine⁹ have been employed for organic NIR fluorophores. Among them, cyanine dyes¹⁰ have attracted the most attention, due to their synthetic accessibility, broad wavelength tunability, and large molar extinction coefficient with moderate fluorescence quantum yields. However, the photostability of the dyes diminishes significantly along with the increase of the π -conjugation system. In particular, the photodegradation is a serious problem for the NIR cyanine dyes having absorbance λ_{max} longer than 700 nm.¹¹ To overcome this limitation, our group has recently developed a photostable NIR cyanine dye library CyNA in which the amine group of CyN was acetylated and thus the photoactivating lone pair electron was removed.¹² As a result, CyNA was much more stable than CyN in general, and CyNA-414 was selected as the most photostable dye from the CyNA library (Scheme 1, upper). While a derivative of CyNA-414 was proved to be superior to currently available commercial NIR dye in *in vivo* mouse imaging,¹³ to further improve the photostability, we designed a new library utilizing the CyNA-414 structure as the key component. Herein we report the synthesis of an ultra-photostable cyanine library (CyR) composed of

80 members (Scheme 1, lower) using a highly efficient and clean solid phase chemistry.

The previous CyNA library was synthesized by solution phase chemistry and the laborious purification step was unavoidable. Inspired by an efficient solid phase chemistry reported for cyanine dyes,¹⁴ we designed a new solid phase route to secure a pure NIR cyanine dye library (CyR) in high speed manner by minimizing a crucial purification step. In order to synthesize the CyR library, firstly, 3-bromopropylamine was loaded to the 2-chlorotrityl resin quite efficiently, and then, a broad range of structural diversity was introduced through a series of primary amines (Chart 1). Here, the nucleophilic displacement of bromide occurred in *N*-methyl-2-pyrrolidone (NMP) solvent under moderate heating conditions, with



Scheme 1 Synthesis of CyR library. Reagents and conditions: (a) DIEA, THF, 3-bromopropylamine, r.t., 12 h; (b) DIEA, RNH₂, NMP, 70 °C, 12 h; (c) HATU, DIEA, r.t., 24 h; (d) 2% TFA in DCM, r.t., 10 min.

^aDepartment of Chemistry & MedChem Program of Life Sciences Institute, National University of Singapore, 117543, Singapore.

E-mail: chmcyt@nus.edu.sg; Fax: +65-6779-1691; Tel: +65-6516-7794

^bCollege of Pharmacy, Sunchon National University, 540-742, South Korea

^cLaboratory of Bioimaging Probe Development, Singapore Bioimaging Consortium, Agency for Science, Technology and Research (A*STAR), 11 Biopolis Way, #02-02 Helios Building, 138667, Singapore

† Electronic Supplementary Information (ESI) available: synthetic procedures and characterization data (NMR, HR-MS) for CyR derivatives; additional photostability data for the CyN, CyNA and CyR compounds and spectral data for CyR library. See DOI: 10.1039/c1ra00498k

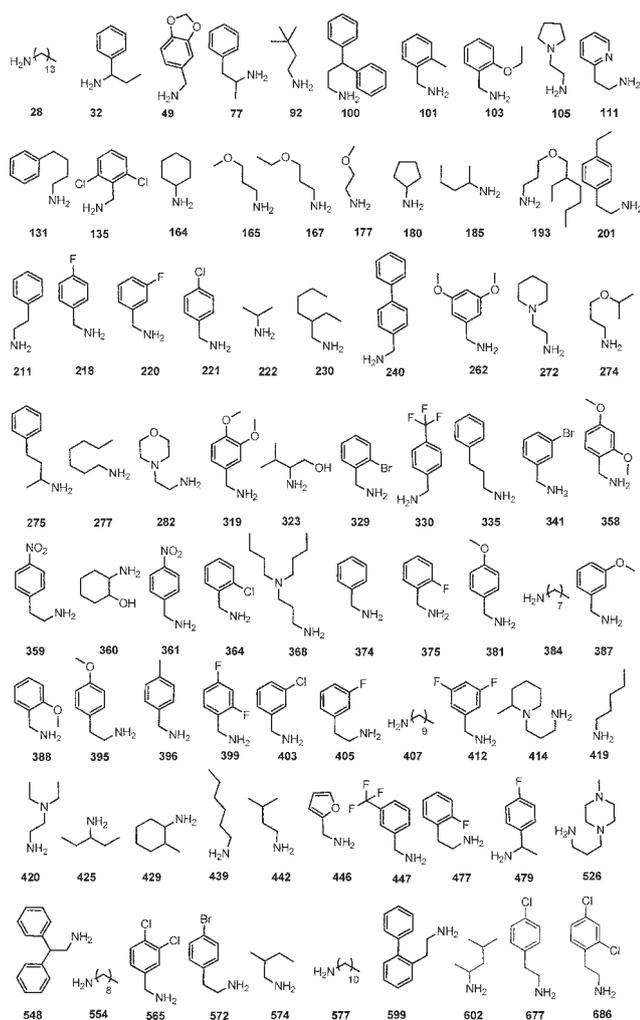


Chart 1 Amine building blocks of the CyR library.

primary amines to lead structurally diversified solid supported secondary amines. The key intermediate **1** (CyNA-414 derivative)¹³ was coupled to this solid phase amine building block by using a standard acid–amine coupling protocol.¹⁵ The final step of acidic cleavage from the solid supported compounds **2** yielded 80 very pure members of the CyR library (average purity is 94% without further purification at 365 nm, Table S1†). It is noteworthy that the current synthetic protocol provides a linker motif with the amino group incorporated in the cyanine scaffold in addition to the structural diversity. The amino linker will be useful for further derivatization of the library compounds with various reporter or affinity tags for bio-conjugation depending on the biological study requirements. The photophysical properties of the 80 members of the CyR library were measured with the maximum absorption wavelengths from 806–807 nm and maximum emission wavelengths varying from 821 nm to 825 nm. The average quantum yield for the CyR compounds is around 10% in DMSO solution (Table S1†).

To evaluate the photostability of CyR in comparison with CyNA and CyN, we examined the decrease of fluorescence intensity of the compounds under a xenon lamp (6 W cm⁻²) for 8 h. The average fluorescence intensity decreases of CyNA and CyN compounds¹² were around 17% and 70% respectively, whereas the newly

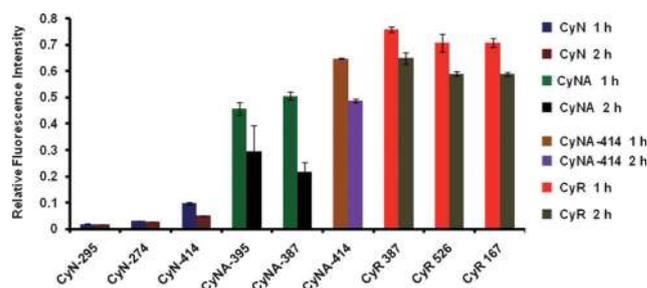


Fig. 1 Photostability assessment under strong UV irradiation in each 10 min interval for 2 h of CyN, CyNA (including CyNA-414) and 3 selected CyR compounds. 1% DMSO in 10 mM HEPES buffer (pH 7.4) was used to dissolve the compounds at a final concentration of 10 μ M. The bar graphs exhibit mean fluorescence intensity after 1 h and 2 h. Values are means \pm s.d. acquired from three experiments.

Table 1 Photodecomposition rate constant of three selected CyR compounds under the strong UV light

Compound	k (s ⁻¹)
CyNA-414	11.16×10^{-5}
CyR 167	8.16×10^{-5}
CyR 387	7.00×10^{-5}
CyR 526	8.33×10^{-5}

synthesized CyR library shows only 5% decrease on average (Table S1) reflecting the superior photostability in general. To differentiate the relative photostability among CyR compounds, we introduced a stronger light condition (UVP Blak-Ray® B-100AP high intensity mercury lamp, 100 W, 365 nm) to all the CyR compounds and monitored the relative fluorescence intensity (F/F_0) at 1 h and 2 h (Table S2†). Three most stable CyR compounds (CyR 167, CyR 387, CyR 526) were selected and further compared with representative CyN and CyNA compounds including CyNA-414 (structures available in Tables S3, S4, S5†) and the results are summarized in Fig. 1. It clearly shows that the three selected CyR compounds exhibit much better photostability compared to CyN and CyNA, even the previous best compound CyNA-414. A further detailed kinetic analysis was performed for the three CyR dyes in comparison with CyNA-414 and the kinetic constants were measured (Table 1, Fig. S1, S2, S3, S4†) showing the superior photostability of new CyR compounds to CyNA-414.

In summary, we designed and demonstrated an efficient solid phase synthesis of an ultra-photostable NIR fluorescence library, CyR. The reported synthetic procedure is quite straightforward and practical to be applied to much bigger library construction with almost no purification step required. Therefore, the CyR library and derivatives might be an extremely useful toolbox to develop novel *in vivo* bioimaging probes.

Acknowledgements

The authors are grateful to the intramural funding from A*STAR (Agency for Science, Technology and Research, Singapore) Biomedical Research Council and the National University of Singapore (NUS) for the financial support (Young Investigator Award: R-143-000-353-123).

References

- (a) V. Ntziachristos, C. H. Tung, C. Bremer and R. Weissleder, *Nat. Med.*, 2002, **8**, 757–760; (b) J. Fabian, H. Kakazumi and M. Matsuoka, *Chem. Rev.*, 1992, **92**, 1197–1226; (c) M. Matsuoka, *Infrared Absorbing Dyes*, Plenum Press, New York, 1990; (d) J. V. Frangioni, *Curr. Opin. Chem. Biol.*, 2003, **7**, 626–634; (e) W. M. Leevy, S. T. Gammon, J. R. Johnson, A. J. Lampkins, H. Jiang, M. Marquez, D. Piwnica-Worms, M. A. Suckow and B. D. Smith, *Bioconjugate Chem.*, 2008, **19**, 686–692; (f) E. M. Sevick-Muraca, J. P. Houston and M. Gurfinkel, *Curr. Opin. Chem. Biol.*, 2002, **6**, 642–650.
- R. B. Mujumdar, L. A. Ernst, S. R. Mujumdar, C. J. Lewis and A. S. Waggoner, *Bioconjugate Chem.*, 1993, **4**, 105–111.
- W. K. Moon, Y. H. Lin, T. O'Loughlin, Y. Tang, D. E. Kim, R. Weissleder and C. H. Tung, *Bioconjugate Chem.*, 2003, **14**, 539–545.
- W. Pham, W. F. Lai, R. Weissleder and C. H. Tung, *Bioconjugate Chem.*, 2003, **14**, 1048–1051.
- D. B. Shealy, M. Lipowska, J. Lipowski, N. Narayanan, S. Sutter, L. Strekowski and G. Patonay, *Anal. Chem.*, 1995, **67**, 247–251.
- K. Jyothish, K. T. Arun and D. Ramaiah, *Org. Lett.*, 2004, **6**, 3965–3968.
- K. Takagi, M. Kawabe, M. Matsuoka and T. Kitao, *Dyes Pigm.*, 1985, **6**, 177–188.
- W. B. Tuemmler and B. S. Wildi, *J. Am. Chem. Soc.*, 1958, **80**, 3772.
- (a) J. Chen, I. R. Corbin, H. Li, W. Cao, J. D. Glickson and G. Zheng, *J. Am. Chem. Soc.*, 2007, **129**, 5798–5799; (b) S. A. Hilderbrand and R. Weissleder, *Curr. Opin. Chem. Biol.*, 2010, **14**, 71–79.
- (a) J. Panda, P. R. Virkler and M. R. Detty, *J. Org. Chem.*, 2003, **68**, 1804–1809; (b) G. A. Reynolds and K. H. Drexhage, *J. Org. Chem.*, 1977, **42**, 885–888; (c) N. Narayanan and G. Patonay, *J. Org. Chem.*, 1995, **60**, 2391–2395; (d) P. J. Sims, A. S. Waggoner, C. H. Wang and J. F. Hoffman, *Biochemistry*, 1974, **13**, 3315–3330; (e) S. J. Mason and S. Balasubramanian, *Org. Lett.*, 2002, **4**, 4261–4264; (f) S. J. Mason, J. L. Hake, J. Nairne, W. J. Cummins and S. Balasubramanian, *J. Org. Chem.*, 2005, **70**, 2939–2949; (g) M. Lopalco, E. N. Koini, J. K. Cho and M. Bradley, *Org. Biomol. Chem.*, 2009, **7**, 856–859.
- M. Lipowska, G. Patonay and L. Strekowski, *Synth. Commun.*, 1993, **23**, 3087–3094.
- A. Samanta, M. Vendrell, R. Das and Y. T. Chang, *Chem. Commun.*, 2010, **46**, 7406–7408.
- A. Samanta, M. Vendrell, S. W. Yun, Z. Guan, Q. H. Xu and Y. T. Chang, *Chem.–Asian J.*, 2011, DOI: 10.1002/asia.201100041.
- M. Lopalco, E. N. Koini, J. K. Choa and M. Bradley, *Org. Biomol. Chem.*, 2009, **7**, 856–859.
- C. Ornelas, R. Lodescar, A. Durandin, J. W. Canary, R. Pennell, L. F. Liebes and M. Weck, *Chem.–Eur. J.*, 2011, **17**, 3619–3629.